A STEREOCONTROLLED TOTAL SYNTHESIS OF A GANGLIO-GANGLIOSIDE GM1b, $IV^{3}NeuAcaGgOse4Cer^{1}$

Mamoru Sugimoto^a), Kazushige Fujikura^a), Shigeki Nunomura^b), Toshio Horisaki^b), Yukishige Ito^b), and Tomoya Ogawa^b)*

a)Central Research Institute, MECT Co., 1780, Kitano, Tokorozawa-shi, Saitama 359 Japan b) RIKEN (The Institute of Physical and Chemical Research), Wako-shi, Saitama, 351-01 Japan

Abstract: A first total synthesis of ganglio ganglioside GM1b, α -D-Neup5Ac-(2 \rightarrow 3)- β -D-Galp-(1 \rightarrow 3)- β -D-GalpNAc-(1 \rightarrow 4)- β -D-Galp-(1 \rightarrow 4)- β -D-Glcp-(1 \rightarrow 1)-Cer, was achieved in a stereocontrolled manner.

In 1973, biotransformation of gangliotetraosylceramide asialo GM1b into GM1b 1 was reported by Yip² and the structure was determined in 1975 by Stoffyn et al³. Subsequently GM1b was isolated from human erythrocyte membranes⁴, rat tumor cells⁵, mouse myeloid leukemia cells⁶, mouse spleen⁷, human brain⁸, and murine T lymphoblast⁹.



As part of our project¹⁰ on the synthesis of *ganglio* gangliosides, we now describe a first total synthesis of GM1b 1, which clearly demonstrates the efficiency of O-2a pivaloyl group in 19b over acetyl group in 19a as an auxiliary¹¹ to achieve coupling between a glycan of *ganglio* series and a ceramide part.

Synthetic plan was delineated in scheme 1. A glycopentaosyl donor 2 armed with an auxiliary at O-2a is designed as a key glycosyl donor along with two kinds of known ceramide derivatives 3^{12} and 4^{13} as key glycosyl acceptors. The donor 2 was disconnected into a glycobiosyl donor 5 and a glycotriosyl acceptor 6. A readily available¹⁴ imidate 7 was chosen as an equivalent to the donor 5. A glycosyl acceptor 6 may be designed as 13ab and prepared as follows. Sn(OTf)2¹⁵ And powdered molecular sieves 4A (MS4A) promoted glycosylation of $9a^{16}$

and 9b¹⁷ with 8¹⁸ in CH₃CN gave 10ab¹⁹ (a 67%; b 72%), which were further converted into glycosyl acceptors 13ab¹⁹ in 4 steps via 11ab¹⁹ and 12ab¹⁹ (1 NH₂NH₂•H₂O in EtOH at 80°, 2 1:10 Ac₂O-MeOH, 3 PhCH(OMe)₂ and TsOH•H₂O in CH₃CN, 4 BH₃NMe₃-AlCl₃ in THF²⁰, a 36%; b 50% overall).





Crucial couplings between 13ab and 7 were achieved in $(CH_2CI_2)_2$ by use of BF₃•OEt₂ as a promotor in the presence of MS4A to afford 14^{19} (a 50%, b 59%) whose configurations at newly introduced glycosidic linkages at C-1d were assigned as β -D by ¹H n.m.r. data after convertion into deblocked derivatives 15ab¹⁹ in 3 steps (1 NaOMe-MeOH, 2 NaOH-aq. MeOH, 3 10% Pd-C and H₂ in 8:2 MeOH-H₂O, a 63%; b 67% overall). Having determined the structures, 14ab were smoothly converted into the key glycopentaosyl donors $19ab^{19}$ in a conventional manner in 4 steps (1 10% Pd-C and H₂ in MeOH, 2 Ac₂O in Py, 3 NH₂NH₂•AcOH in DMF for 5 min at 50°²¹, 4 Cl₃CCN and DBU²² in (CH₂Cl)₂, a 36%; b 62%).



Having necessary key intermediates 3, 4 and 19ab in our hands, the coupling experiments were performed employing BF3.0Et2 in the presence of MS4A at 20° under Ar. Reaction of the donor 19a with the acceptor 3 in CHCl3 and 4 in $(CH_2CI)_2$ afforded 15.4 and 17.4% yield of the desired product 20a and 21a, respectively. On the other hand, the donor 19b armed with pivaloyl auxiliary at O-2a upon reaction with the acceptor 3 did afford a 32% yield of the coupled

product 20b, showing clearly the efficacy of pivaloyl over acetyl auxiliary at O-2a in the coupling between a ceramide equivalent and a ganglio series glycosyl donor. The result is in harmony with our previous experiments on $lacto^{23}$ and $globo^{24}$ series glycosyl donors. Both of the fully protected glycosphingolipids 20ab were converted into the target molecule GM1b 1 in 2 steps (1 NaOMe in 1:1 MeOH-THF, 2 NaOH in 1:1:1 MeOH-THF-H₂O, finally purified by Sephadex LH-20 in 12:6:1 CHCl₃-MeOH-H₂O a and b: 78%).

In summary, a first total synthesis of GM1b was achieved in a stereocontrolled manner. ¹H-N.m.r. data recorded in 49:1 (CD3)2SO-D2O for both synthetic 1 and human brain⁸ GM1b were found in complete agreement.

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Reference and Notes

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- 19 Physical data for key compounds are given below. Values of $[\alpha]_D$ and $\delta_{H,C}$ were recorded for

solutions in CHCl₃ and CDCl₃, respectively, at $23\pm3^\circ$, unless noted otherwise. 1: δ H (49:1 (CD3)2SO-D2O, 24°) 4.513 (d, 7.0 Hz, 1c), 4.232 (d, 8.1 Hz, 1d), 4.191 (d, 7.7 Hz, 1b), 4.159 (d, 7.7 Hz, 1a), 2.771 (dd, 4.0, 12.5 Hz, 3eeq), 1.888 and 1.794 (2 s, 2NAc); 10a: $[\alpha]_D$ +2.6° (c 1.1); δ_H 6.149 (dd, 3.3 and 11.4 Hz, 3c), 5.554 (d, 3.7 Hz, 4c), 5.381 (d, 8.4 Hz, 1c); δ_C 102.6 (1a), 102.1 (1b), 99.8 (1c), and 51.6 (2c); 10b: [a]D -12.5° (c 2.0); $\delta_{\rm H}$ 6.157 (dd, 3.3 and 11.4 Hz, 3c), 5.558 (d, 3.3 Hz, 4c), 5.340 (d, 8.4 Hz, 1c), 1.219 (s, Bu^t); δC 102.2 (1b), 99.8 (1a) and 99.8 (1c), 51.6 (2c); 11a: [α]D +10.3° (c 1.0); $\delta_{\rm H}$ 1.478 (s, Ac); $\delta_{\rm C}$ 103.0 (1c), 102.6 (1a), 102.4 (1b), 62.5 (6c), 56.5 (2c); 11b: [α]D -9.7° (c 1.4); $\delta_{\rm H}$ 5.119 (dd, 7.7 and 9.2 Hz, 2a), 1.499 (s, Ac), 1.165 (s, Bu^t); 12a: [α]D +24.5° (c 1.0); $\delta_{\rm H}$ 5.595 (s, CHPh), 1.612 (s, Ac); $\delta_{\rm C}$ 103.0 (1c), 102.6 (1ab), 101.3 (CHPh), 55.2 (2c); 12b; [α]D +1.2° (c 1.2); $\delta_{\rm H}$ 5.583 (s, CHPh), 5.113 (dd, 8.1 and 9.2 Hz, 2a), 4.475 (d, 8.1 Hz, 1a), 4.482 (d, 7.7 Hz) and 4.428 (d, 7.3 Hz, 1bc); δC 103.1 (1c), 102.9 (1b), 101.4 (CHPh), 99.9 (1a), 55.2 (2c); 13a: $[\alpha]_D$ +7.7° (c 1.2); δ_H 1.524 (s, Ac); δ_C 102.8, 102.6 and 102.4 (1abc), 56.4 (2c); 13b: $[\alpha]_D$ -9.4° (c 0.7); δ_H 5.130 (dd, 7.7 and 9.2 Hz, 2a), 4.489 (d, 7.7 Hz, 1a), 1.537 (s, Ac), 1.164 (s, tBu); δ_C 102.8 and 102.5 (1bc), 99.9 (1a), 56.4 (2c); 14a: $[\alpha]_D$ +16.6° (c 2.3); δ_H 7.40-7.15 (m, 8Ph), 3.837 (s, OMe); 14b: $[\alpha]_D$ +6.6° (c 0.5); δ_H 7.40-7.15 (m, 7Ph), 3.841 (s, OMe); 15a: δ_H (D₂O, TMSCD₂CO₂CO₂Na, 24°) 5.227 (d, 0.5 H, 4.0 Hz, 1aa), 4.714 and 4.709 (2d, 8.4 Hz, 1caβ), 4.675 (d, 0.5 H, 8.1 Hz, 1ab), 4.526 (d, 7.7 Hz, 1d), 4.494 (d, 8.1 H, 1b), 2.761 (dd, 4.8 and 12.8 Hz, 3eeq), 2.051, 2.049, and 2.037 (3s, 3Ac), 1.797 (t, 12.1 Hz, 3eax), 15b: δ_H (D₂O, ^tBuOH 24° and 60°+) 5.315⁺ (d, 0.5 H, 3.7 Hz, 1aa), 4.824* (d, 0.5 H, 8.1 Hz, 1ab), 4.731* and 4.718* (2 d, 8.0 Hz, 1cab), 4.691* (dd, 0.5 H, 3.7 and 10.3 Hz, 2aa), 4.655* (dd, 0.5 H, 8.4 and 10.3 Hz, 2ab), 4.504 (d, 7.7 Hz, 1d), 4.450 and 4.440 (2 d, 7.7 Hz, 1bαβ), 2.739 (dd, 4.8 and 12.8 Hz, 3eeq), 2.031, 2.027, and 2.016 (3 s, 3Ac), 1.776 (t, 12.1 Hz, 3eax); 16a: [α]D +6.2° (c 2.7, MeOH); δ_H (CD3OD) 3.858 (s, OMe), 2.575 (dd, 4.5 and 12.2 Hz, 3eeq); 16b: $[\alpha]_D$ +2.0° (c 1.0, MeOH); δ_H (CD3OD) 3.864 (s, OMe), 2.585 (dd, 4.8 and 12.7 Hz, 3eeq), 1.544 (t, 12.1 Hz, 3eax), 1.228 (s, tBu); 17a (α : β =1:1): δ H 6.274 (d, 0.5 H, 3.7 Hz, 1a α), 5.665 (d, 0.5 H, 8.4 Hz, 1a β), 3.841 (s, OMe), 2.585 (dd, 4.6 and 12.0 Hz, 3eeq); 17b (α : β =1:1): δ H 6.308 (d, 0.5 H, 3.7 Hz, 1aa), 5.703 (d, 0.5 H, 8.4 Hz, 1ab), 3.842 (s, OMe), 2.577 (dd, 4.7 and 12.5 Hz, 3eeq), 1.701 (t, 12.5 Hz, 3eax); 18a: $\delta_{\rm H}$ 3.842 (s, OMe), 2.578 (dd, 4.8 and 12.8 Hz, 3eeq); 18b: $\delta_{\rm H}$ 3.840 (s, OMe), 2.578 (dd, 4.7 and 12.2 Hz, 3eeq); 19a: $[\alpha]_D$ +31.0° (c 1.3); δ_H 6.499 (d, 3.7 Hz, 1a), 3.843 (s, OMe), 2.579 (dd, 4.5 and 12.4 Hz, 3eeq); δC 100.7, 100.0, 98.2, 96.8, 92.9 (5 anomeric C), 90.7 (CCl₃), and 37.6 (3e); 19b: $[\alpha]_D$ +31.0° (c 2.5); δ_H 2.580 (dd, 4.8 and 12.8 Hz, 3eeq); 20a: $[\alpha]_D$ +15.4° (c 0.3); δH 8.008 (d, 2 H, 8.4 Hz, Ph), 7.548 (t, H, 8.1 Hz, Ph), 7.439 (t, 2 H, 8.1 Hz, Ph), 5.869 (dt, 15.4 and 7.0 Hz, 5cer), 4.694 (d, 7.7 Hz, 1d), 4.427 (d, 7.7 Hz, 1a), 4.372 (d, 8.0 Hz, 1b), 2.578 (dd, 4.8 and 12.5 Hz, 3eeq), 1.694 (t, 12.5 Hz, 3eax), 0.879 (t, 7.0 Hz, 2CH₂CH₃); SC 100.5, 100.5, 100.1, 98.2, 96.9 (5 anomeric C); 20b: δH 2.577 (dd, 4.4 and 12.8 Hz, 3eeq), 0.880 (t, 6.6 Hz, 2CH₂CH₃); 21a: [α]D +4.8° (c 0.3); $\delta_{\rm H}$ (HOHAHA) 7.7-7.3 (m, aromatic), 5.269 (dd, 7.7 and 14.3 Hz, 4cer), 4.688 (d, 7.7 Hz, 1d), 4.411 (d, 8.1 Hz, 1a), 4.375 (d, 7.7 Hz, 1b), 3.843 (s, OMe), 2.579 (dd, 4.8 and 12.8 Hz, 3eeq), 1.005 (s, tBu), 0.883 (t, 7.0 Hz, CH₂CH₃), 0.879 (t, 7.0 Hz, CH₂CH₃).

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